

MK

06/13/13

1. Cadmium detector
2. Arsenic detector #1
3. Arsenic detector #2
4. C2R1-A
5. C2R1-B
6. K08104 #1
7. I13461 #1
8. 008 #1
9. 008 #2
10. 009 #1

Mix

Buffer	2.5	* 11 =	27.5	✓
MgCl2	1.5		16.5	✓
dNTP	1		11	✓
VR	.5		5.5	✓
VF2	.5		5.5	✓
Tag	.5		5.5	✓
dH ₂ O	13.5		148.5	✓
	<u>20</u>			

6/13

- grew liquid culture of DHS α

- grew plates of:

- K824008

- I13401

- DHS α

- K824009

- K081014

- K824012

- K174015

- ran PCR of:

1 - Cadmium detector

2 - Arsenic detector #1

3 - Arsenic detector #2

4 - C2R1 - A

5 - C2R1 - B

6 - K081014 #1

7 - I13461 #1

8 - K824008 #1

9 - K824008 #2

10 - K824009 #1

- did plasmid preps of:

- K824008

- I13401

- I13401

- K824009

- K081014

- K824012

- K174015

- did a digest of:

- Cd Detector #1

- Lead Detector #2

- Arsenic Detector #3

- Arsenic Detector #4

- C2R1 - A

- K081014

- Arsenic Detector 2

- Arsenic Detector

- I13401

- C2R1 - B

- Cadmium Detector

• Procedure for digest

- 10 λ DNA	} x12 =	24 λ Buffer 2	-
- 2 λ Buffer 2		12 λ Eco-R1	
- 1 λ Eco-R1		12 λ PST-1	
- 1 λ Pst-1		72 λ dH ₂ O	-
- 6 λ dH ₂ O			↳ master mix
<u>20 λ digest</u>			

- distribute 10 λ of master mix to each tube and incubate

• Made liquid cultures of:

- Nitrate Detector (Precons) A
- Nitrate Detector (Precons) C
- Nitrate Detector (Precons) D
- 504450 (Chlor)
- 504450 Amp [sample from 6-21-11]
- 504450 Amp [sample 2-22-10]