# Protocols #XXX : Study of M15 [pRep4;pQE30::KR] cell response to light illumination iGEM Grenoble-EMSE-LSU

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## **Materials and reagents:**

- M9 minimum medium (with iron sulfate and elementary traces)
- M15[pRep4;pQE30::KR] cells
- Sterile LB medium
- Sterile LB-Agar
- Ampicillin (100μg/mL), Kanamycin (50μg/mL)
- IPTG (0.05M)
- 1.5 mL Eppendorf tubes
- 50 mL Falcon tubes
- 90 mm Petri dishes
- Incubator
- LED Lamp (specifications)
- Spectrophotometer (for fluorescence and OD610 measurements)
- 3 Autoclaved 100 mL Erlenmeyers
- Aluminum fold
- Spreader
- Black 96-well plates
- Disposable absorbance cuvettes
- Pipettes, with sterile pipet tips

### **Protocol**

## Day 1 (in the evening):

- 1) Pre culture M15[pRep4;pQE30::KR] cells ON (37°C, 200 rpm) in a 50 mL Falcon tube, filled with 10mL M9 medium, supplemented with 200μg/μL Ampicillin and 50μg/μL Kanamycin.
- 2) Wrap the 3 Erlenmeyers in aluminum fold
- Prepare 100-150 LB-Agar plates, supplemented with 200μg/μL Ampicillin and 50μg/μL Kanamycin

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### Day 2:

- 1) Supplement 100 mL M9 minimum medium with iron sulfate, elementary traces, 200µg/µL Ampicillin, 50µg/µL Kanamycin and 0.05mM IPTG.
- 2) Pre warm the mix at 37°C
- 3) Measure the OD610 of your M15[pRep4;pQE30::KR] pre culture, and re suspend it at 0.015 in each of the Erlenmeyer, filled with pre warmed and complete M9 medium.
- 4) Immediately measure OD610 and fluorescence (540/630 nm). Spread 50μL of diluted cell samples on agar plates, using 10<sup>4</sup>, 10<sup>5</sup> and 10<sup>6</sup> dilutions (made in sterile LB medium). Incubate the resulting plates ON (37°C)

Note: Let agar plates open under a sterile environment for 30 min before performing the plating, to dry them up.

- 5) Repeat step 4) every 30min-1h for 3 hours.
- 6) At t = 180 min, unwrap one of the Erlenmeyer, and start illuminating it in the incubator with the LED lamp.
- 7) Repeat step 4) every 30-1h for 8 hours.

#### Day 3:

- 8) Count the number of colonies on each of the agar plates (discard those displaying more than 300 colonies or less than 30 colonies)
- 9) Calculate the number of living cells per µL using the following formula

#cells/µL = (#colonies\*dilution)/50