## Protocols \#XXX : Study of M15 [pRep4;pQE30::KR] cell response to light illumination iGEM Grenoble-EMSE-LSU

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## Materials and reagents:

- M9 minimum medium (with iron sulfate and elementary traces)
- M15[pRep4;pQE30::KR] cells
- Sterile LB medium
- Sterile LB-Agar
- Ampicillin $(100 \mu \mathrm{~g} / \mathrm{mL})$, Kanamycin $(50 \mu \mathrm{~g} / \mathrm{mL})$
- IPTG (0.05M)
- 1.5 mL Eppendorf tubes
- 50 mL Falcon tubes
- 90 mm Petri dishes
- Incubator
- LED Lamp (specifications)
- Spectrophotometer (for fluorescence and OD610 measurements)
- 3 Autoclaved 100 mL Erlenmeyers
- Aluminum fold
- Spreader
- Black 96-well plates
- Disposable absorbance cuvettes
- Pipettes, with sterile pipet tips


## Protocol

Day 1 (in the evening):

1) Pre culture $\mathrm{M} 15\left[\mathrm{pRep} 4 ; \mathrm{pQE} 30:\right.$ KR] cells $\mathrm{ON}\left(37^{\circ} \mathrm{C}, 200 \mathrm{rpm}\right)$ in a 50 mL Falcon tube, filled with 10 mL M9 medium, supplemented with $200 \mu \mathrm{~g} / \mu \mathrm{L}$ Ampicillin and $50 \mu \mathrm{~g} / \mu \mathrm{L}$ Kanamycin.
2) Wrap the 3 Erlenmeyers in aluminum fold
3) Prepare 100-150 LB-Agar plates, supplemented with $200 \mu \mathrm{~g} / \mu \mathrm{L}$ Ampicillin and $50 \mu \mathrm{~g} / \mu \mathrm{L}$ Kanamycin

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Day 2:

1) Supplement 100 mL M9 minimum medium with iron sulfate, elementary traces, $200 \mu \mathrm{~g} / \mu \mathrm{L}$ Ampicillin, $50 \mu \mathrm{~g} / \mu \mathrm{L}$ Kanamycin and 0.05 mM IPTG.
2) Pre warm the mix at $37^{\circ} \mathrm{C}$
3) Measure the OD610 of your M15[pRep4;pQE30::KR] pre culture, and re suspend it at 0.015 in each of the Erlenmeyer, filled with pre warmed and complete M9 medium.
4) Immediately measure OD610 and fluorescence ( $540 / 630 \mathrm{~nm}$ ). Spread $50 \mu \mathrm{~L}$ of diluted cell samples on agar plates, using $10^{4}, 10^{5}$ and $10^{6}$ dilutions (made in sterile LB medium). Incubate the resulting plates $\mathrm{ON}\left(37^{\circ} \mathrm{C}\right)$

## Note: Let agar plates open under a sterile environment for 30 min before performing

 the plating, to dry them up.5) Repeat step 4) every $30 \mathrm{~min}-1 \mathrm{~h}$ for 3 hours.
6) At $\mathrm{t}=180 \mathrm{~min}$, unwrap one of the Erlenmeyer, and start illuminating it in the incubator with the LED lamp.
7) Repeat step 4) every $30-1 \mathrm{~h}$ for 8 hours.

## Day 3:

8) Count the number of colonies on each of the agar plates (discard those displaying more than 300 colonies or less than 30 colonies)
9) Calculate the number of living cells per $\mu \mathrm{L}$ using the following formula
\#cells/ $\mu \mathrm{L}=(\# c o l o n i e s * d i l u t i o n) / 50$
