Gel Extraction of DNA		Prepare Gel	Load DNA	Separate DNA	Insert Filter Paper & Dialysis Tubing	Collect DNA
		< 45 min.	< 5 min.	< 45 min.	< 5 min.	< 10 min.
75mL	1x TAE buffer	Add TAE, agarose, heat to dissolve. Add stain; pour into mold w/ 8-lane comb.	Add 5 µL dye to 25µL digested DNA (500ng). Load 5µL ladder, 30µL DNA/dye samples. Leave at least one lane between samples.	Run @ 120 V until fully resolved. Meanwhile, cut filter paper and dialysis tubing slightly wider and taller than comb teeth. Puncture a hole in the micro-centrifuge tube with a syringe and place it inside Eppendorf tube.	Make an incision just downstream of separated DNA band. Insert filter paper and dialysis tubing such that filter paper is between DNA and dialysis tubing. (Filter paper absorbs the DNA, dialysis tubing inhibits migration through the gel). Visualize to ensure proper alignment.	Run @ 120 V for 5 min to capture DNA. Place filter paper and dialysis tubing into micro-centrifuge tube. Visualize to ensure no (or very little) residual DNA left in gel. Spin @ 13,200 rpm for 30 s to collect DNA in Eppendorf tube. Proceed to ligation or store @ -20° C.
0.525g - 0.75g	Agarose					
0.75µL	1000x SYBR Safe Stain					
25µL	Digested DNA (20ng/µL)					
5µL	6x loading dye					
1ct.	1.5mL Eppendorf tube					
1ct.	0.65mL micro-centrifuge tube					
	Syringe					
	Sharp razor blade					
	Tweezers					
	Filter paper					
	Dialysis tubing					