# BBa\_K1123019

### **Basic Information**

<u>Basic Function</u>: The expression of POLY(ARGININE-SERINE) <u>Description</u>: This part holds the DNA information for the POLY(ARGININE-SERINE) protein, so that it can be placed Behind a promoter and expressed.

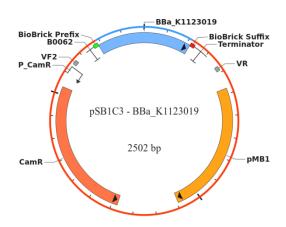


Related Parts: BBa\_K1123007
Authors: Ardjan van der Linden
Data Collection: Ardjan van der Linden

<u>Affiliation</u>: TU-Eindhoven <u>Team</u>: TU-Eindhoven 2013

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<u>Date Submitted</u>: 31-08-2013 <u>Date Updated</u>: 31-08-2013 <u>Biosafety</u>: Risk Group 1 <u>Availability</u>: Unavailable <u>Sequence</u>: Confirmed



<u>Additional Comments</u>: This Brick has been tested in the pET28a vector and not the pSB3C1. All experiments were performed in this vector. The part has been tested using this vector to check the protein expression, the protein itself and if expressed properly will then be used functionally in the brick BBa\_K1123008

### Reference:

### **Device Information**

<u>Device Name</u>: BBa K1123019 <u>Assembly</u>: BioBrick

<u>Device Type</u>: Coding <u>Protocol</u>: -<u>Description</u>: Protein sequence <u>Scars</u>: No

<u>Components</u>: none <u>Insertion</u>: Plasmid

Vector: pET28a

<u>Additional Comments</u>: The DNA of this part was not submitted to the registry. It was therefore not transferred to the pSB1C3 vector. The biobrick was therefore only tested in the pET28a vector.

#### Reference:

## **Plate Imaging**

### **BASIC INFORMATION**

<u>Purpose</u>: Control of ligation into pET28a vector

Chassis: E.coli

Strain: NB (NovaBlue)

**Protocols:** Standard plating procedure

Date: 24/07/13

### **GROWTH CONDITIONS**

<u>Media Type</u>: Agar + Kanamycin <u>Antibiotic</u>: Kanamycin (30μL/mL)

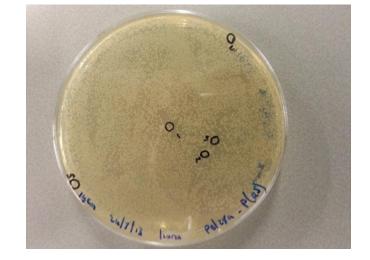
<u>Vessel</u>: Petri dish <u>Incubation</u>: 37°C <u>Growth Time</u>: 15 hours

### **MEASUREMENT INFORMATION**

Data Type: Picture

<u>Location</u>: TU-Eindhoven BioLab Machine Name: Digital Camera

Reporter Used: none



Additional Comments: The growth of colonies was seen simply by photographing the growth plates after

incubation. Reference: None

## **Restriction Mapping**

#### **BASIC INFORMATION**

Purpose: Check ligation and PCR results, also used to check

protein sequence length.

Chassis: E.coli

Strain: NB (NovaBlue)

**Protocols:** Standard gel electroforesis

Date: 08/08/19

#### **MEASUREMENT INFORMATION**

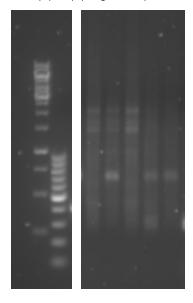
<u>Data Type</u>: Gel electrophoresis Location: TU-Eindhoven BioLab

Machine Name: N/A

Enzymes Used: PCR primers
Total Time: 60 minutes
Voltage Used: 100 V

Ladder Used: New England Biolans 100 bp ladder

Ladders
Kbp | 100bp | Digested samples



<u>Additional Comments</u>: We performed colony PCR on the plates shown above using T7 FW and RW primers These samples were then run on gel giving an indication of correct ligation into pET28a. From this gel we can see that almost none of the constructs had been ligated and digested correctly. However upon continuation this was not the case, the samples 2 and 5 were correct.

Reference: none

## **Growth Curve**

### **BASIC INFORMATION**

<u>Purpose</u>: To prepare for protein expression.

<u>Chassis</u>: E.coli <u>Strain</u>: BL21

**Protocols**: Standard Expression protocol

Date:

### **GROWTH CONDITIONS**

Media Type: LB + Kanamycin

<u>Vessel</u>: culture flask <u>Volume</u>: 400mL

Incubation: 37°C, 250rpm

### **MEASUREMENT INFORMATION**

<u>Data Type</u>: Growth Curve (OD vs Time)

Location: TU-Eindhoven Biolab

Machine Name: N/A Time Interval: N/A Total Time: N/A

Additional Comments: The expression of this protein has been tested, unfortunately no data is available.

Reference: None

## **Future Work**

This brick will later be used in a composite part, BBa\_K1123007, once in that part, the same protein will be expressed anaerobically. We will need to then perform the same controls on the composite part to show that the expression remains equal, aerobically as well as anaerobically.