BBa_K1123013

Basic Information

<u>Basic Function</u>: The expression of Protamine-1-Optimized <u>Description</u>: This part holds the DNA information for the Protamine-1-Optimized protein, so that it can be placed Behind a promoter and expressed.

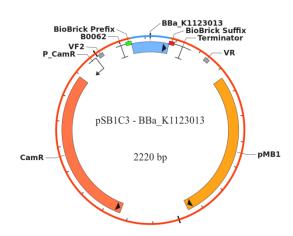


Related Parts: BBa_K1123013
Authors: Ardjan van der Linden
Data Collection: Ardjan van der Linden

<u>Affiliation</u>: TU-Eindhoven Team: TU-Eindhoven 2013

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<u>Date Submitted</u>: 31-08-2013 <u>Date Updated</u>: 31-08-2013 <u>Biosafety</u>: Risk Group 1 <u>Availability</u>: Unavailable <u>Sequence</u>: Confirmed



<u>Additional Comments</u>: This Brick has been tested in the pET28a vector and not the pSB3C1. All experiments were performed in this vector. The part has been tested using this vector to check the protein expression, the protein itself and if expressed properly will then be used functionally in the brick BBa K1123002

Reference:

Device Information

<u>Device Name</u>: BBa_K1123013 <u>Assembly</u>: BioBrick

<u>Device Type</u>: Coding <u>Protocol</u>: -<u>Description</u>: Protein sequence <u>Scars</u>: No

<u>Components</u>: none <u>Insertion</u>: Plasmid

Vector: pET28a

<u>Additional Comments</u>: The DNA of this part was not submitted to the registry. It was therefore not transferred to the pSB1C3 vector. The biobrick was therefore only tested in the pET28a vector.

Reference:

Plate Imaging

BASIC INFORMATION

Purpose: Control of ligation into pET28a vector

Chassis: E.coli

Strain: NB (NovaBlue)

Protocols: Standard plating procedure

Date: 24/07/2013

GROWTH CONDITIONS

<u>Media Type</u>: Agar + Kanamycin <u>Antibiotic</u>: Kanamycin (30μL/mL)

<u>Vessel</u>: Petri dish <u>Incubation</u>: 37°C <u>Growth Time</u>: 15 hours

MEASUREMENT INFORMATION

Data Type: Picture

<u>Location</u>: TU-Eindhoven BioLab <u>Machine Name</u>: Digital Camera

Reporter Used: none

Additional Comments: The growth of colonies was seen simply by photographing the growth plates after

incubation.

Reference: None



BASIC INFORMATION

Purpose: Check ligation and PCR results, also used to check

protein sequence length.

Chassis: E.coli

Strain: NB (NovaBlue)

Protocols: Standard gel electroforesis

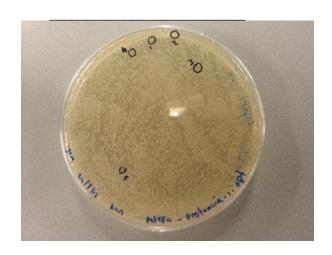
Date:08/08/13

MEASUREMENT INFORMATION

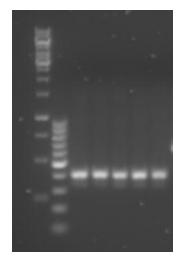
<u>Data Type</u>: Gel electrophoresis <u>Location</u>: TU-Eindhoven BioLab

Machine Name: N/A

<u>Enzymes Used</u>: PCR primers Total Time: 60 minutes



Ladders Kbp | 100bp | Digested samples



Voltage Used: 100 V

Ladder Used: New England Biolans 100 bp ladder

Additional Comments: We performed colony PCR on the plates shown above using T7 FW and RW primers

These samples were then run on gel giving an indication of correct ligation into pET28a.

Reference: none

Growth Curve

BASIC INFORMATION

<u>Purpose</u>: To prepare for protein expression.

<u>Chassis</u>: E.coli <u>Strain</u>: BL21

Protocols: Standard Expression protocol

Date:

GROWTH CONDITIONS

Media Type: LB + Kanamycin

<u>Vessel</u>: culture flask <u>Volume</u>: 400mL

Incubation: 37°C, 250rpm

MEASUREMENT INFORMATION

<u>Data Type</u>: Growth Curve (OD vs Time)

Location: TU-Eindhoven Biolab

Machine Name: N/A
Time Interval: N/A
Total Time: N/A

Additional Comments: Thei protein has been expressed properly, unfortunately no data is available for the

cell growth.

Reference: None

Future Work

This brick will later be used in a composite part, BBa_K1123001, once in that part, the same protein will be expressed anaerobically. We will need to then perform the same controls on the composite part to show that the expression remains equal, aerobically as well as anaerobically.